

International Journal of Nutrition and Agriculture Research

Journal home page: www.ijnar.com



DETERMINATION OF MICRO- AND MACRO-ELEMENT CONCENTRATIONS AND THEIR DIVERSITY FROM THE COMMON BEAN (*PHASEOLUS VULGARIS*, L) LANDRACES OF ZAMBIA

Alex Abaca*¹, Rod Scott¹, Robert Lungu², Kennedy K. Muimui², John R. Beeching¹

¹*Department of Biology and Biochemistry, University of Bath, Bath, BA2 7AY, United Kingdom and Abi Zonal Agricultural Research and Development Institute (Abi ZARDI).

²Misamfu Regional Research Centre, Zambia Agricultural Research Institute (ZARI), P.O.Box.410055, Kasama, Zambia.

ABSTRACT

Bio-fortification of crops is one of those cheaper options to managed micronutrient deficiencies (MND), compared to supplementation and food fortification. For major crops, bio-fortification relies on the natural sources of variation in the mineral contents of the available genotypes including the landraces. Therefore, the main objective of this research was to determine the mineral content variation amongst the landraces and commercial varieties of common bean from Zambia alongside the CIAT reference lines using the Atomic Absorption Spectrophotometer (AAS) method. We observed significant differences ($p < 0.05$) among the different sup-populations of the landraces, commercial varieties and CIAT reference lines for copper, sodium, iron, zinc, manganese, potassium, calcium and magnesium. On average, a 2-fold higher concentration was observed for the landraces compared to the commercial varieties and the CIAT reference lines for some minerals. Strong positive correlations ($p < 0.05$) were revealed amongst these minerals. The sup-populations of these landraces with particularly high mineral contents for iron and zinc were identified and these remain very useful in the Zambian common bean improvement programmes for food and nutritional security.

KEYWORDS

Common bean, Landraces, Bio-fortification, Macro and Micro elements, Zambia.

Author for Correspondence:

Alex Abaca,
Department of Biology and Biochemistry,
University of Bath, Bath, BA2 7AY, United Kingdom.
Abi Zonal Agricultural Research and Development
Institute (Abi ZARDI).

Email: abal-2007@hotmail.com

INTRODUCTION

Common bean (*Phaseolus vulgaris*, L) is grown for its fresh leaves and pods as vegetables, and dry grains in Latin America, Eastern and Southern Africa and South East Asia¹⁻⁵, with excellent nutritional properties linked to its high protein, carbohydrate, vitamin and mineral content. Despite the contribution from common bean and other food

staples, the prevalence of micronutrient deficiencies (MND) is still high in developing countries and is caused mainly by lack of essential vitamins (Vitamin A) and minerals (Iron and Zinc)⁶. Iron deficiency anaemia (IDA) has been reported to be the most prevalent micronutrient condition globally^{7,8}. Specifically, these authors presented that 65.5% of pre-school children suffering from anaemia, while 45.7% - 48.2% of women of reproductive age suffer from IDA, and that Vitamin A deficiency (VAD) affects 190 million children under the age of five. A total of 49 known essential nutrients needed to sustain human life together with some of their average energy allowance (AEA), recommended dietary allowance (RDA), estimated safe and adequate daily dietary intakes (ESADDI), minimum requirement (MR) and both the enhancing and decreasing anti-nutrient substances that promote/inhibit micronutrient bioavailability⁹.

The roles of these minerals in the human diet have been elaborated upon by many authors¹⁰⁻¹². For instance, calcium (Ca) and magnesium (Mg) play important roles in the development of bone and structural tissue formation, glucose and protein absorption and metabolism, regulation and dilation of blood vessels and regular heart beat^{10,12}. The deficiency in Ca and Mg causes weak bone and structural connecting tissue formation, hypertension, and poor glucose absorption¹³. Ca deficiency is linked to some chronic diseases such as *osteoporosis*, and that Mg deficiency leads to energy production faltering and insufficient production¹¹. Iron (Fe) is a crucial component of haem proteins, haemoglobin, and myoglobin required for oxygen transportation and vascular functions¹⁴, zinc (Zn) serves as a cofactor in many enzymatic reactions¹⁵, copper (Cu) is a coenzyme and a crucial cofactor in Fe utilisation and is required for cytochrome oxidase redox chemical reaction¹⁶, manganese (Mn) is essential for immune system and effective food metabolism in addition to serving as cofactors in many enzymatic reactions¹⁷.

Due to the prevalence of these MNDs, several approaches have been put in place to mitigate them through supplementation and food fortification. However, supplementation and fortification are cost

ineffective and do not reach the rural poor¹⁸ hence the development of bio-fortification. Bio-fortification has been defined differently but the key components are development of micronutrient-dense staple crops using the best traditional breeding practices and modern biotechnology¹⁹ or improving the nutritional content of staple crops by breeding varieties that have a high content of the three limiting micronutrients (Vitamin A, Iron, Zinc) or their precursors than conventional ones²⁰, and a number of advantages associated with it are being advanced^{21,22}. Iron content variations have been observed to occur between genotypes and wild and cultivated beans, although the wild beans had only a narrow advantage in iron content over cultivated ones^{1,5}. The wide genetic variations amongst genotypes in iron and zinc contents in Tanzania between leaves (310.49 ppm of Fe, and 28.03 ppm of Zn) and seeds (55.01 ppm of Fe, and 31.4 ppm of Zn) has been reported².

Specific studies that rely on the mineral composition based on germplasm from African origin are few, as seen in the work of Typhone and Nchimbi-Msolla² and Mukamuhirwa *et al*³ for Tanzania and Uganda respectively. This lack of information on the available common bean genotypes hampers a cheap source of food supplementation through bio-fortification of common beans for East and Southern African countries. The mineral composition and / or content of bean germplasm from major producing countries in these regions, including Zambia, remain unknown. Furthermore, how these minerals vary between landraces and commercial varieties has not been reported anywhere to the best of our knowledge.

Therefore, the main objective of this study to determine and present the macro- and micro-element concentrations of the sub-populations within the Zambian common bean landraces, and to compare how these vary with respect to the defined CIAT reference lines and Zambian commercial varieties. This is aimed at encouraging bio-fortification through breeding by identifying potential parental breeding lines among the sub-populations of these landraces with high iron and zinc content for food security and nutritional breeding programmes.

MATERIAL AND METHODS

The plant materials

The sub-populations within the four Zambian landraces of Lusaka yellow, Lundazi, Mbala mixture and Solwezi; the CIAT control genotypes of 'Diacol Calima' (G4494, B and C) for the Andean gene pool, and 'ICA Pijao' (G5773) for the Mesoamerican gene pool; and the Zambian commercial varieties as detailed here under (Table No.1) were used for this study. The landraces were collected by the Zambian Agricultural Research Institute's (ZARI) breeders, CIAT reference lines were sourced from the CIAT, Colombia, gene bank, and the commercial lines were collected from the Lusuntha border market between Zambia and Malawi. The seeds were kept in cool dry conditions and planted in the tropical glass house at the University of Bath under the following conditions: on a mixed compost of coarse, medium and fine in the ratio of 2:2:1 in a 5 litres pots that was supplemented with a slow releasing nutrients Osmocote Extract (Standard 12-14M, ICL, UK) at the rate of 5 grams per pot, Temperature of 22–28°C, relative humidity (RH) of 40–80%, and a light period of 14 hours per day. These plants were maintained to maturity and their seeds harvested for this study. A total of 50 samples were used for this study, that is, Lusaka yellow had 8 sub-populations, Lundazi had 8 sub-populations, Mbala mixture had 7 sub-populations, and Solwezi had 8 sub-populations. Additionally, there were sub populations that overlapped between landraces: 1 overlap for Lusaka yellow and Mbala mixture, 2 for Lundazi, Mbala mixture, and Solwezi, and 8 for Mbala mixture and Solwezi, totalling to 42 sub-populations from all the landraces. In addition to these landrace sub-populations, there were four Zambian-Malawian varieties of Katwetwe, Kabulangeti, Sugar beans, and White bean that were collected from the Lusuntha border market between the two countries due to their market dominance, and finally the 4 CIAT reference varieties of G5773, G4494A, G4494C, G14470) were included in this study.

Sample preparation and Acid digestion

The 100-seed weight of each of the 50 sub-population was determined prior to sample preparation as indicated in Table No.1 above. Six

grams of sample were ground into a fine powder using the Andrew James wet and dry grinder (Andrew James, UK Ltd). Two grams of finely ground bean flour were stored in boiling tubes for subsequent acid digestion. Acid digestion was conducted as described by^{2,12}. Briefly, to the fine ground bean flour in each of the boiling tube was added 20 ml of 6M 70% nitric acid for trace element analysis (Sigma-Aldrich, UK) and allowed to stand for an overnight. The boiling tubes were then arranged vertically in a glass beaker and placed in a Heraeus B6 Incubator (Fisher Scientific, UK) at a temperature 50°C for 3 hours. The digested samples were cooled to room temperature, filtered through Whatman filter paper, and diluted with trace element grade deionised water (Sigma-Aldrich, UK) in a 25 ml volumetric flask. The solution was thus ready for determination of macro and micro elements in the atomic absorption spectrophotometer (AAS) method of Perkin-Elmer 2380 (Fisher Scientific, UK).

Determination of micro and macro elements in the samples

Calcium (Ca), magnesium (Mg), copper (Cu), potassium (K), manganese (Mn), phosphorus (P), iron (Fe) and zinc (Zn) were quantified using Atomic Absorption Spectroscopy (AAS) method of Perkin-Elmer 2380 (Fisher Scientific, UK), and their concentrations were converted and expressed in mg100g⁻¹ from the absorbance read in the (AAS) as described in^{11,23,24}. For Iron and Zinc, the readings were evaluated against the standard curves prepared from the Iron diluted to a concentration of 100 mg l⁻¹ and Zinc diluted to 50 mg l⁻¹ as explained by²⁵.

Data analysis

Data were collected in triplicate and the averages were computed in Microsoft Excel 2013. One-way ANOVA was applied to evaluate the variance of these micro and macro elements parameters; the Pearson coefficient was used to verify the existence of statistically significant correlations among the variables; the multivariate analysis of main PCA components was performed, with the aim to detect the existence of clusters grouping amongst the bean accessions according to their mineral concentrations. All these analyses were run using PAST software, Version 3.16²⁶ and R Software²⁷.

RESULTS

Micro- and macro-nutrient variation in common-bean landraces

A significant difference ($p < 0.05$) was recorded for the 100-seed weight (100SW), and micro and macro nutrients in common bean landraces from Zambia, CIAT reference lines and the Zambian commercial varieties (Table No.2). Specifically, the landrace of Lundazi was highest in calcium, a sub-population that overlaps between Lusaka yellow and Mbala mixture (LY+MM) was highest in iron, manganese, and zinc, a sub-population that overlaps between Mbala mixture and Solwezi (MM+SO) was highest in copper, potassium and magnesium, and the CIAT line (G4494B) was highest in sodium. It is important to point here that 8 of the 9 highest values for the 100SW and macro- and micro-nutrients in common bean were from the Zambian landraces whereas 4 of the lowest values were from the Zambian commercial varieties, 2 from CIAT lines and 3 from the landraces.

Micro and macro nutrient variation in common bean based on populations

Considering population averages as a whole, Lusaka yellow had three of the highest values for iron, potassium and calcium; Mbala mixture had two of the highest values for manganese and potassium; G5773 from CIAT lines had the highest values for copper and zinc; G4494B from a CIAT line had the highest values for sodium and magnesium; and the commercial variety of Kabangeti had the highest value for seed weight (Table No.3). The landraces of Lundazi and Solwezi had neither the lowest nor highest values for the micro and macro nutrients. The six of the eight lowest values all came from the Zambian commercial varieties while the other two came from the CIAT lines. Thus, there were high values for these minerals in the landraces followed by CIAT lines and lower values were recorded among the Zambian commercial varieties. The Zambian landraces had 5 of the 8 highest values on average for copper, manganese, zinc, potassium, and calcium, CIAT reference lines had 3 of the 8 for iron, sodium and manganese, and the commercial varieties were high in 100 seed weight only. The Zambian commercial varieties had 7 of the 8 lowest

average values for copper, iron, manganese, zinc, potassium, magnesium and calcium in common bean, and the other only lowest value from the landraces was for Sodium. At population and sub-population levels, analysis of variance showed there were significant differences ($p < 0.05$) between these mineral contents among populations. Mann-Whitney pairwise significance analysis also showed that there were significant differences ($p < 0.05$) between any pair of these micro and macro nutrients in the common bean germplasm studied. However, there was no correlation between any of the mineral content and 100SW.

Micro and macro nutrient variation in common bean based on seed colour

When the seeds were classified according to seed colour there was no statistical difference ($p < 0.05$) between the macro and micro mineral contents for the common bean germplasm from Zambia. For the results presented here, there was a very minor separation between dark red and maroon, and they could be treated interchangeably. 100 seed weight was highest in green and lowest in yellow; copper was highest in maroon and lowest in white; Iron was highest in maroon and lowest in white; manganese was highest in yellow and lowest in pink; sodium was highest in red and lowest in white; zinc was highest in maroon and lowest in white; potassium was highest in red and lowest in purple; magnesium was highest in red and lowest in green; and calcium was highest in red and lowest in green (Table No.4). Red and maroon took most high values for the mineral concentrations measured, and white took most of lowest values for these minerals, with green colour taking two and purple and pink taking one each of these low values of the mineral concentration. Black, brown, and grey did not take any of the highest or lowest values and were all close to the top higher values.

Pair wise correlation analyses of the micro and macro nutrients in common bean

Pairwise correlation analyses produced strong significant ($p < 0.05$) positive relationships between some of the minerals in common beans germplasm from Zambia (Table No.5). These strong positive significant relationships were observed between

copper and iron and zinc; iron and manganese and zinc; sodium and copper and iron; potassium and magnesium and calcium; magnesium and calcium; and calcium and sodium. Frequently, strong positive correlations occurred between micro-nutrients themselves and between macro-nutrients themselves and less frequently between a pair involving micro- and macro-elements relationships. The negative significant correlations were observed between sodium and zinc and calcium, whereas the strongest positive significant correlation was observed between Sodium and Copper.

Principal component analysis (PCA) and Neighbor joining clustering based on the mineral concentrations in common beans

The multivariate analysis using principal component analysis (PCoA) generated the partial distribution of all the sub-populations based on the micro and macro element concentrations (Figures No.1). The results indicated that the macro and micro element concentrations can be explained by 8 axes, of which only 4 had significant contributions to the spatial separation of the sub-populations explaining 70.54 % of the total variability. The first axis explained 22.85% of the total variability with elements of Cu, Fe, Na, Zn, K, and Ca being positively significant; the second axis explained 18.41% with all the elements being positively significant except K; the third axis explained 16.53% with Cu, Fe, Mg, and Ca being positively significant; and the fourth axis explained 12.76% with all the elements except Ca and Fe being significant, with the eigenvalues of 1.96, 1.58, 1.42, and 1.10 respectively. However, there were no clear clustering patterns for all the sub-populations based on the macro and micro mineral concentrations in the neighbour joining.

DISCUSSION

The main objective of this study was to determine the micro and macro nutrient concentrations in dry seeds of common bean germplasm from Zambia. This study focused on the micro nutrients in common beans as these have been reported to cause the highest global health risks, particularly iron and zinc deficiencies^{7,9}. Among the macro elements, sodium, calcium, potassium and magnesium were

included because of their nutritional importance in the human diet as major components of bones and teeth, and proper functioning of muscles and central nervous system²⁴.

The results from this study showed that, there are significant differences ($p < 0.05$) in the concentrations of these micro and macro elements in common bean landraces from Zambia. These results agree with previous studies for example, the variations of iron and zinc between leaves and seeds of Tanzanian common bean genotypes, and plant parts² was reported. They showed that from leaves, the average values were 310.49 ppm of Fe, and 28.03 ppm of Zn whereas from seeds were 55.01 ppm (5.5 mg) of Fe, and 31.4 ppm (3.14 mg) of Zn. The average Fe and Zn concentration of 5.5 mg/100g and 3.5 mg/100g respectively in the core collection of CIAT genotypes¹ was presented. The work of²⁴ on the germplasm of Madeira Island of Portugal and showed that on average the mineral concentrations were 1890 mg/100g for K, 150 mg/100g for Mg, 6.01 mg/100g for Fe, 1.01 mg/100g for Cu, 3.01 mg/100g for Zn, and 1.45 mg/100g for Mn. The mineral contents of common bean genotypes from India were found to be on average as 1.81 mg/100g Fe, 0.78 mg/100g Zn and 20.30% protein content. The mineral concentrations in the 29 genotypes from USA were studied and reported that Zn ranged from 3.4 to 6.4 mg/100g and Fe was from 0.89 to 11.29 mg/100g by¹¹.

It is apparent from the above that there has been no specific study directed towards the mineral concentrations in the landraces of common bean from many major bean-growing regions of the world. This could be the probable reason why the values of the micro and macro elements reported in this study are higher than those that had previously been reported. However, several factors have been suggested that effect the mineral concentrations of common beans: common bean plant parts²; growing environment and genotypes variations^{11,23}; origin, genotypes, environmental conditions (temperature, soils, and fertilization), growing conditions²⁴; and the weeding regimes that affect both mineral nutrient uptake and retention in the plant as well as soil moisture for zinc²⁹. When the results of this study

are broken down in to landraces, CIAT lines and Zambian commercial lines (Table No.3) there are higher values on average for landraces, followed by CIAT lines and lowest in the commercial varieties. It is important to note further here that, the averages of the mineral content of the Zambian commercial varieties fall within most of the values reported earlier. This implies that, the presence of landraces in this study is the reason for the higher values than the average values for mineral content observed, and it demonstrates how important these landraces can be used in improving nutritional component of commercial varieties already in production.

Akond¹¹ studied the mineral compositions of USA genotypes and identified 7 genotypes with high iron and zinc concentrations which were recommended to be used as parental material for mineral content breeding in USA. In this current study we identified 23 sub-populations with iron concentrations above average (21.38 mg/100g): 6 in Lusaka yellow, 3 in Lundazi, 3 in Mbala mixture, 6 in Solwezi, one in an overlap between Lusaka yellow and Mbala mixture, 3 in an overlap between Mbala mixture and Solwezi and 2 in CIAT lines of G5573 and G4494C. This study further identified 26 sub-populations with zinc concentration above average (8.08 mg/100g): 7 in Lusaka yellow, 4 in Lundazi, 4 in Mbala mixture, 4 in Solwezi, one in an overlap between Lusaka yellow and Mbala mixture, 5 in overlap between Mbala mixture and Solwezi, and 2 in CIAT lines of G5573 and G4494C. It is important to point out here that the CIAT line G4494 had two plant types: a dwarf type with white flowers and a semi climber with a pink flower colour; therefore, G4494C refers to the latter plant type. CIAT line G4494B can also be considered for future breeding work since it had the highest values for sodium. The overlap between Lusaka yellow and Mbala mixture, mentioned above, is the dominant composition in Lusaka yellow, and had the highest average values for Fe, Zn and Mn with elongate/kidney seed shape with dark blue colour around its helium. This could partly explain why the landrace of Lusaka yellow beans have a higher preference in the markets within Lusaka and other parts of Zambia.

Additional result from this study showed that there was no direct correlation ($p < 0.05$) between mineral contents and 100SW. This agrees with the findings of Akond¹¹ and Maraghan²³ when they reported a similar result from their earlier studies. Therefore, seed size is mainly an attribute for seed yields as explained by Agung³⁰ in Faba beans (*Vicia faba* L) and not for mineral content estimation. However, this study disagrees with the finding of Beebe¹ that Fe content tends to be present at higher values in the Andean gene pool than the Mesoamerican gene pool. Here, we report Fe in CIAT G5773 – Mesoamerican at 25.0 mg/100g, and in CIAT G4494C – Andean at 24.5 mg/100g that agrees with the results of Akond¹¹ where they also observe no direct relation with the gene pools in relation to the mineral contents for the genotypes from USA.

Positive and negative correlations between the micro and macro elements have been reported in earlier studies^{1, 2, 11, 12, 24, 28}. This study also reported positive and significant correlations ($p < 0.05$) between micro elements as presented in Table No.5. The strong and significant correlations between micro and macro elements have been explained to mean that the genetic factors for increasing one mineral content co-segregate with the genetic factor for increasing the other mineral content with which they share a significant correlation^{2,9}. Therefore, deducing from these results, increasing the content of iron would increase the content for zinc, manganese, and sodium while increasing the zinc content would increase the content for iron, copper, and sodium. A similar observation was made for macro elements such that increasing the potassium content would increase the content for magnesium and calcium, increasing calcium would increase potassium, sodium and magnesium, and increasing magnesium would increase calcium and potassium.

To explore further the observed correlations among the mineral contents in common bean for breeding purposes, a focus paper by Welch and Graham⁹ observed that there is significant variability to increase the concentrations of iron and zinc, and that the traits required for the genetic improvement of iron and zinc concentrations are stable across different bean growing environments, although their

concentrations are affected by GXE interactions. Beebe¹ had confirmed that, there is an environmental and seasonal stability in the iron concentrations for the CIAT collections. The genetic parameters of iron and zinc concentrations in Andean bean seeds by looking at their concentrations from crosses between two sets parents (IAC Boreal x Light Red Kidney and Ouro Branco x Light Red Kidney), their F₁ plants, F₁ reciprocals, F₂ plants, F₂ reciprocals, and backcrosses (BC₁₁ and BC₁₂) were studied³¹. Zemolin and the co-authors³¹ concluded that there are no maternal effects, and that the seeds of the F₁ generation will represent fertilisation between the parents in both hybrids combinations. Therefore, these results mean that there is no time that needs to be wasted in undertaking reciprocal crosses with F₁ plants before backcrossing hence allowing segregating populations of common beans with high mineral contents to be generated and deployed within a shorter period of time. On the other hand, although Possobom *et al*³² observed a significant expression of maternal effects in the Mesoamerican genepool, they linked this expression of maternal effects to iron accumulation in the seed coat in Mesoamerica beans or seed embryo in Andean beans.

Possobom and his colleagues³² then suggested that the selection of superior common bean recombinants for iron concentration should begin at F₃ generation if iron accumulates more in the seed coat (Mesoamerican) and at F₂ if the iron accumulates more in the seed embryo (Andean).

Finally, on the breeding and inheritance of seed iron and zinc concentrations in common bean, for example, 6 QTLs for zinc and 5 QTLs for iron that are clustered on the upper half of the linkage group B11 were identified²⁵. Other QTLs for Zn were identified on linkage groups B3, B6, B7 and B9, and B4, B6, B7 and B8 for Fe. This information means that the scientists are getting so close to identifying the candidate gene(s) for Fe and Zn concentrations in common bean, which will be very useful in the era of marker assisted breeding that will shorten the breeding cycle by allowing screening for these minerals at seedling stage of growth for common bean.

Table No.1: Details of the sub-populations of common bean with their seed coat colour and 100-seed weight (100SW) used for the macro and micro mineral concentration determination

S.No	Landrace	Sub-population	Seed Colour	100SW
1	Lusaka Yellow (LY)	LY1	Yellow	27.05
		LY2	Creamy yellow	29.47
		LY3	Deep yellow	34.52
		LY4	Olive green	48.00
		LY5	Yellow	31.26
		LY6	Creamy yellow	24.31
		LY7	Brownish yellow	29.51
		LY8	Yellow	26.91
2	Lundazi (LU)	LU1	Dark red	42.54
		LU2	Maroon	20.75
		LU3	Red	22.51
		LU4	Dark brown	33.58
		LU5	Black	24.27
		LU6	Purple	41.17
		LU7	Red mottled	27.94
		LU8	Purple	25.19

3	Mbala Mixture (MM)	MM1	Pinkish mottled	52.68
		MM2	Purplish mottled	30.67
		MM3	Grey to brownish green	38.29
		MM4	Yellow	31.45
		MM5	Yellow	26.90
		MM6	Pinkish with yellow speckles	50.15
		MM7	Brownish yellow	30.44
4	Solwezi (SO)	SO1	Pinkish mottled	51.94
		SO2	Purple mottled	32.16
		SO3	Dark Red/Maroon	40.16
		SO4	Pink	20.55
		SO5	Red	30.01
		SO6	Red mottled	37.40
		SO7	Pink with black mottled	38.50
		SO8	Pink with black speckles	30.07
5	LY and MM overlap	LYMM	Yellow	37.51
6	LU, MM and SO overlap	LMS1	Purplish with speckles	39.21
		LMS2	Grey to brownish green	29.23
7	MM and SO overlap	MS1	White	31.83
		MS2	Pinkish mottled	31.57
		MS3	Purplish mottled	29.41
		MS4	Brownish to Pinkish mottled	27.02
		MS5	Dark grey with speckles	37.72
		MS6	Blackish with brown mottling	38.52
		MS7	Dark brownish mottled	49.74
		MS8	Black with Cream mottling	33.16
8	CIAT Reference Lines	G5773	Black	19.76
		G4494B	Red mottled	39.70
		G4494C	Red mottled	31.89
		G14470	Cream with purple stripped	36.91
9	Zambian/Malawian commercial Lines	Long White	White with purple dotted helium	36.35
		Kabulangeti	Dark grey speckled	46.16
		Katwetwe	Purple mottled	30.81
		Sugar Bean	Cream purple stripped	37.56

Table No.2: Variations in the mineral content: copper (Cu), iron (Fe), manganese (Mn), sodium (Na), zinc (Zn), potassium (K), magnesium (Mg), and calcium (Ca) in the landraces of Lusaka yellow (LY), Lundazi (LU), Mbala mixture (MM), and Solwezi (SO), four CIAT Reference Lines, and four Zambian commercial Varieties

Populations	Sub-Pops.	100SW	Cu	Fe	Mn	Na	Zn	K	Mg	Ca
Lusaka Yellow	LY1	27.05	2.40	21.50	3.50	0.60	7.75	3900.00	570.00	650.00
	LY2	29.47	2.50	22.00	3.00	0.70	7.75	5100.00	590.00	950.00
	LY3	34.52	2.20	26.00	3.50	3.40	9.25	4700.00	540.00	800.00
	LY4	48.00	2.55	21.50	3.00	1.00	8.25	4500.00	480.00	375.00
	LY5	31.26	1.50	18.50	3.00	0.65	8.50	4800.00	650.00	675.00
	LY6	24.31	2.95	28.50	4.50	1.20	9.75	5700.00	650.00	775.00
	LY7	29.51	2.80	31.00	4.00	0.70	10.00	5800.00	590.00	650.00
	LY8	26.91	2.10	23.50	4.00	1.70	9.25	5900.00	600.00	950.00
Lundazi	LU1	42.54	2.65	18.50	3.50	1.85	9.00	5000.00	540.00	600.00
	LU2	20.75	3.00	29.50	3.50	0.75	9.50	5200.00	620.00	550.00
	LU3	22.51	2.50	25.50	3.50	5.35	7.00	5300.00	620.00	525.00
	LU4	33.58	2.50	23.00	3.50	0.90	9.00	5200.00	600.00	600.00
	LU5	24.27	2.10	19.50	2.50	1.05	6.00	5000.00	670.00	525.00
	LU6	41.17	2.20	20.50	3.50	4.20	6.75	5000.00	510.00	550.00
	LU7	27.94	2.10	17.50	4.00	2.30	8.75	5200.00	600.00	1275.00
	LU8	25.19	1.80	16.50	3.50	0.80	6.50	4400.00	620.00	875.00
Mbala Mixture	MM1	52.68	2.40	26.50	2.75	0.80	9.00	5900.00	640.00	725.00
	MM2	30.67	2.10	20.00	2.50	0.50	8.25	5200.00	550.00	600.00
	MM3	38.29	1.75	21.00	2.00	0.45	10.00	4900.00	540.00	650.00
	MM4	31.45	2.45	21.50	3.00	0.75	7.25	5400.00	590.00	900.00
	MM5	26.90	1.80	19.50	3.00	0.60	6.50	3600.00	480.00	575.00
	MM6	50.15	0.85*	9.50	2.00	1.40	5.50	3900.00	460.00	225.00
	MM7	30.44	2.65	25.50	2.50	0.80	8.50	4400.00	480.00	425.00
Solwezi	SO1	51.94	2.55	21.50	2.50	0.90	9.25	4000.00	510.00	575.00
	SO2	32.16	2.65	25.00	3.50	0.70	7.00	4300.00	510.00	500.00
	SO3	40.16	2.90	24.00	3.25	0.90	11.25	4500.00	550.00	400.00
	SO4	20.55	2.50	26.50	3.00	0.90	8.00	4400.00	550.00	450.00
	SO5	30.01	2.15	19.00	2.50	1.20	6.50	5000.00	630.00	450.00
	SO6	37.40	2.30	17.50	2.25*	0.65	7.00	4800.00	510.00	425.00
	SO7	38.50	2.30	23.00	2.50	1.10	9.00	4900.00	510.00	400.00
	SO8	30.07	2.40	26.50	3.50	0.85	8.75	4800.00	590.00	525.00
LY + MM	LYMM	37.51	2.65	34.00	4.50	1.35	12.00	5000.00	540.00	925.00
LU+MM+SO	LMS1	39.21	1.60	16.00	2.50	0.60	6.50	4400.00	510.00	425.00
	LMS2	29.23	2.35	19.50	3.50	0.70	7.00	5400.00	550.00	725.00
MM + SO	MS1	31.83	2.15	17.00	2.50	0.60	6.50	5600.00	650.00	700.00
	MS2	31.57	2.25	21.00	2.75	0.60	8.25	4700.00	600.00	475.00
	MS3	29.41	2.50	27.50	4.00	0.90	9.50	6100.00	690.00	725.00
	MS4	27.02	2.05	7.00*	2.50	0.90	7.00	6000.00	620.00	650.00
	MS5	37.72	2.75	22.00	3.50	0.70	9.25	5700.00	610.00	950.00
	MS6	38.52	3.10	29.50	3.50	0.80	10.50	5500.00	620.00	875.00
	MS7	49.74	3.15	27.00	3.00	0.55	10.50	4400.00	510.00	625.00
	MS8	33.16	2.40	20.50	2.50	0.70	8.00	4700.00	570.00	650.00

CIAT Lines	G5773	19.76*	2.50	25.00	4.00	2.50	9.50	5000.00	540.00	575.00
	G4494B	39.70	2.45	19.50	3.50	9.30	7.25	5900.00	680.00	450.00
	G4494C	31.89	2.30	24.50	3.50	0.75	8.25	5500.00	680.00	625.00
	G14470	36.91	2.15	20.00	2.50	0.50*	7.25	430.00	490.00	300.00
Zambian Commercial Lines	Long White	36.35	1.60	14.00	3.00	0.80	6.50	3500.00*	500.00	250.00*
	Kabulangeti	46.16	1.65	13.00	2.75	2.90	5.50	3600.00	430.00	400.00
	Katwetwe	30.81	1.35	11.00	3.00	1.50	6.50	3600.00	380.00*	325.00
	Sugar Bean	37.56	1.10	11.50	3.50	0.75	4.00*	3600.00	440.00	325.00
Mean		33.89	2.27	21.38	3.15	1.34	8.09	4786.60	563.20	602.50
Std Err		1.15	0.07	0.79	0.09	0.21	0.23	132.51	10.00	29.79
Coeff of var		24.01	21.40	25.99	19.28	111.99	19.76	19.58	12.55	35.17

*lowest values, the bold values are the highest values for each mineral concentration in mg/100g and 100SW

Table No.3: Variation in the mineral content by populations: copper (Cu), iron (Fe), manganese (Mn), sodium (Na), zinc (Zn), potassium (K), magnesium (Mg), and calcium (Ca) among the Zambian Landraces, CIAT reference lines and Zambian Commercial varieties

Origin	Populations	No of sub-pops	100SW	Cu	Fe	Mn	Na	Zn	K	Mg	Ca
Zambian Landraces	Lusaka Yellow	9.00	32.06	2.41	25.17**	3.67	1.26	9.17	5044.44**	578.89	750.00**
	Std Error		2.40	0.14	1.70	0.20	0.30	0.45	18.14	18.14	62.36
	Lundazi	10.00	30.64	2.28	20.60	3.35	1.85	7.60	5010.00	584.00	665.00
	Std Error		2.53	0.41	1.35	0.15	0.52	0.41	110.00	16.94	78.28
	Mbala Mixture	18.00	35.86	2.28	21.36	9.92**	0.76	8.33	5044.44**	567.22	656.94
	Std Error		1.87	0.13	1.56	0.16	0.06	0.41	170.70	15.29	44.96
	Solwezi	18.00	34.90	2.45	21.67	2.96	0.79	8.32	4955.56	571.67	584.72
	Std Error		1.81	0.09	1.26	0.12	0.04	0.36	146.02	13.41	38.73
Mean for Landraces		33.36	2.36 ^b	22.20	4.98 ^b	1.17 ^a	8.36 ^b	5013.61 ^b	575.45	664.17 ^b	
Standard Error		2.15	0.19	1.47	0.16	0.23	0.41	111.22	15.95	56.08	
CIAT Reference Lines	G5773	1.00	19.76*	2.50**	25.00	4.00	2.50	9.50**	5000.00	540.00	575.00
	G4494B	1.00	39.70	2.45	19.50	3.50	9.30**	7.25	5900.00	680.00**	450.00
	G4494C	1.00	31.89	2.30	24.50	3.50	0.75	8.25	5500.00	680.00**	625.00
	G14470	1.00	36.91	2.15	20.00	2.50*	0.50*	7.25	430.00	490.00	300.00
	Mean		32.07 ^a	2.35	22.25 ^b	3.38	3.26 ^b	8.06	4207.50	597.50 ^b	487.50
	Standard Error		4.41	0.08	1.45	0.31	2.06	0.53	1272.55	48.71	72.53
Zambian Commercial Varieties	Long White	1.00	36.35	1.60	14.00	3.00	0.80	6.50	3500.00*	500.00	250.00*
	Kabulangeti	1.00	46.16**	1.65	13.00	2.75	2.90	5.50	3600.00	430.00	400.00
	Katwetwe	1.00	30.81	1.35	11.00*	3.00	1.50	6.50	3600.00	380.00*	325.00
	Sugar Bean	1.00	37.56	1.10*	11.50	3.50	0.75	4.00*	3600.00	440.00	325.00
	Mean		37.72 ^b	1.43 ^a	12.38 ^a	3.06 ^a	1.49	5.63 ^a	3575.00 ^a	437.50 ^a	325.00 ^a
	Standard Error		3.17	0.13	0.69	0.16	0.50	0.59	25.00	24.62	30.62

*lowest population value, **highest population value, ^alowest mean value, ^bhighest mean value, and all concentrations were reported in mg/100g

Table No.4: Variation in the mineral contents by seed colours: Zambian landraces, CIAT lines and Zambian commercial varieties based on seed colours. The number in bracket after seed colour represents the number of sub-populations in that seed colour

Seed Colour	Mean/Std Err	100SW	Cu	Fe	Mn	Na	Zn	K	Mg	Ca
Black (5)	Mean	30.84	2.48	23.50	3.00	1.23	8.60	5020.00	582.00	605.00
	Standard Error	3.80	0.17	1.78	0.32	0.32	0.76	131.91	28.53	78.82
Brown (3)	Mean	36.78	2.57	19.00	3.00	0.78	8.83	5200.00	576.67	625.00
	Standard Error	6.75	0.32	6.11	0.29	0.11	1.01	461.88	33.83	14.43
Green (1)	Mean	48.00	2.55	21.50	3.00	1.00	8.25	4500.00	480.00*	375.00*
Grey (4)	Mean	37.85	2.13	18.88	2.94	1.19	7.94	4900.00	532.50	681.25
	Standard Error	3.46	0.26	2.02	0.36	0.57	1.03	463.68	37.50	113.36
Maroon (2)	Mean	30.46	2.95	26.75	3.38	0.83	10.38	4850.00	585.00	475.00
	Standard Error	9.71	0.05	2.75	0.13	0.08	0.88	350.00	35.00	75.00
Pink (7)	Mean	38.23	2.15	21.64	2.71*	0.85	8.14	4700.00	557.14	510.71
	Standard Error	4.93	0.22	2.30	0.18	0.12	0.47	263.67	22.64	58.72
Purple (8)	Mean	34.05	1.92	18.50	3.25	1.24	6.75	3978.75*	518.75	503.13
	Standard Error	1.95	0.19	2.09	0.19	0.44	0.53	580.66	34.35	73.25
Red (07)	Mean	33.14	2.35	20.29	3.25	3.06	7.68	5242.86	608.57	621.43
	Standard Error	2.68	0.07	1.25	0.24	1.20	0.38	139.48	24.54	112.78
White (2)	Mean	34.09	1.88*	15.50*	2.75	0.70*	6.50*	4550.00	575.00	475.00
	Standard Error	2.26	0.28	1.50	0.25	0.10	0.00	1050.00	75.00	225.00
Yellow (11)	Mean	29.94*	2.36	24.68	3.50	1.13	8.77	4936.36	570.91	752.27
	Standard Error	1.13	0.13	1.47	0.20	0.25	0.46	228.14	17.29	52.05

*lowest value, bold is the highest value for the mineral content in mg/100g.

Table No.5: Pair wise correlations coefficients for the mineral content of Copper (Cu), Iron (Fe), Manganese (Mn), Sodium (Na), Zinc (Zn), Potassium (K), Magnesium (Mg), and Calcium (Ca) in common beans

S.No		Cu	Fe	Mn	Na	Zn	K	Mg	Ca
1	Cu		0.000	0.003	0.927*	0.000	0.007	0.006	0.017
2	Fe	0.785*		0.000	0.866*	0.000	0.026	0.013	0.016
3	Mn	0.403	0.518*		0.091	0.003	0.016	0.039	0.000
4	Na	0.013	-0.025 ^a	0.242		0.422	0.199	0.381	0.720*
5	Zn	0.699*	0.764*	0.407	-0.116 ^a		0.013	0.053	0.002
6	K	0.380	0.315	0.338	0.185	0.350		0.000	0.000
7	Mg	0.381	0.350	0.291	0.127	0.275	0.674*		0.000
8	Ca	0.335	0.338	0.489	-0.052 ^a	0.431	0.535*	0.509*	

*strong positive significant correlations, and ^anegative and significant correlations (p < 0.05)

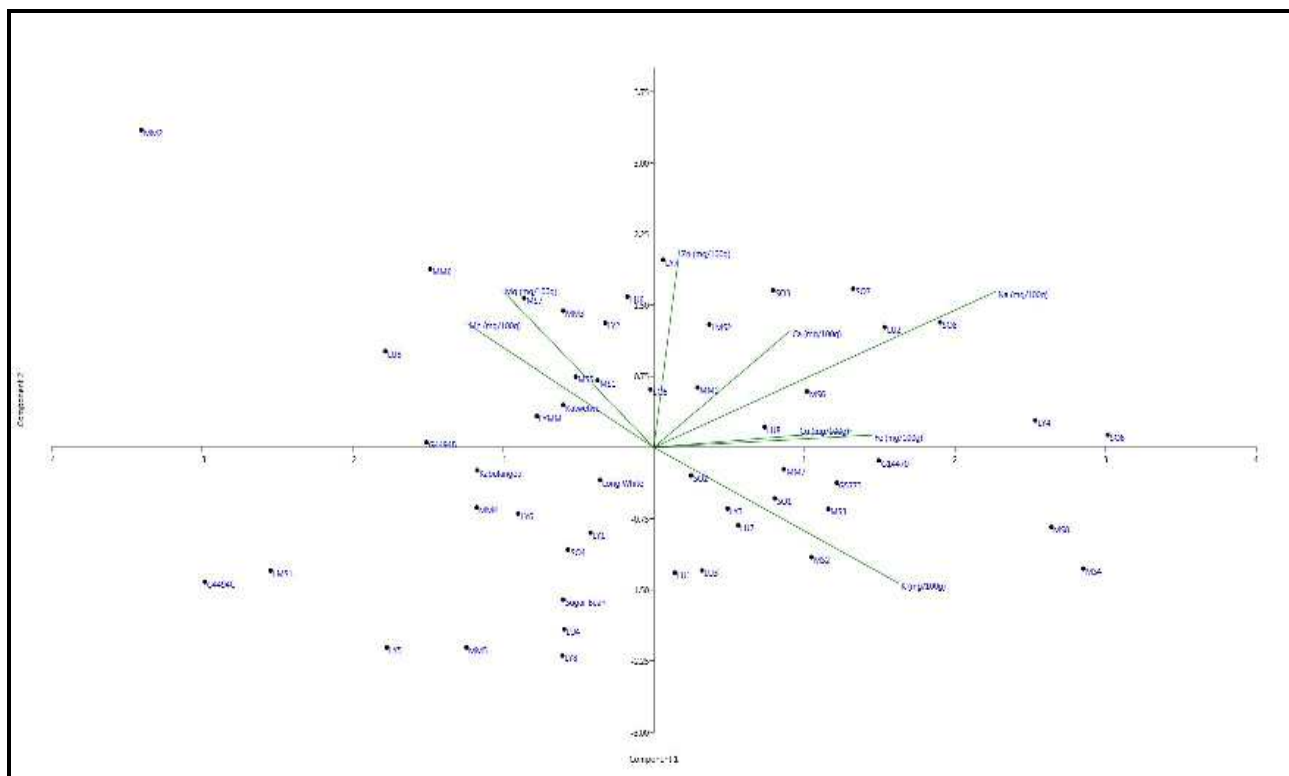


Figure No.1: Representation of the Euclidean bi-plot by principal component analysis (PCA) with transformed data for all the variables in the analysis. Legend: Ca – calcium; Cu – Copper; Fe – Iron; K – Potassium; Mg – Magnesium; Mn- Manganese; Na – Sodium; and Zn – Zinc

CONCLUSION

The PCoA distributions result in this research supports the initial assumption that there is a wide natural variation in the concentrations of macro and micro elements among these common bean landraces from Zambia that can be exploited for common bean improvement programmes in Zambia, and or provide for a short term material exchange between Zambia and her neighbouring/regional countries for common bean breeding programmes in Southern and Eastern Africa. The identified sub-populations with high amount of iron and zinc present an opportunity to breed new varieties of common bean with enhanced iron and zinc content based on the available common bean germplasm within Zambia. Participatory common bean breeding of these identified sub-populations of landraces and commercial varieties within Zambia and the neighbouring countries and evaluating their progenies for iron and zinc contents remain the next course of action towards food and nutritional security using common beans.

ACKNOWLEDGEMENT

The authors acknowledged and thanked the financial support from the University of Bath, Crops-Innovations (<http://crop-innovations.org>) and Self-Help Africa (<https://selfhelpafrica.org/uk/>) to undertake this research. The authors further acknowledged the dedication from the glasshouse technicians James and Robert in looking after the plants, and Alan Carver from the department of Chemistry who prepared standards and conducted mineral-content determinations.

CONFLICT OF INTEREST

We declare that we have no conflict of interest.

BIBLIOGRAPHY

1. Beebe S, Gonzalez A V, Rengifo J. Research on trace minerals in the common bean. *Food and Nutrition Bulletin*, 21(4), 2000, 387-391.
2. Tryphone G M, Nchimbi-Msolola S. Diversity of common bean (*Phaseolus vulgaris* L.)

- genotypes in iron and zinc contents under greenhouse conditions, *African Journal of Agricultural Research*, 5(8), 2010, 738-747.
3. Mukamuhirwa F, Tusiime G, Mukankusi C, Gibson P, Edema R. Potential sources of high iron and zinc content in Ugandan bean germplasm, *In: Third RUFORUM Biennial Meeting*, 2012, 24-28.
 4. Petry N, Boy E, Wirth J P, Hurrell R F. The potential of the common bean (*Phaseolus vulgaris*) as a vehicle for iron biofortification, *Nutrients*, 7(2), 2015, 1144-1173.
 5. Chavez-Mendoza C, Sanchez E. Bioactive compounds from Mexican varieties of the common bean (*Phaseolus vulgaris*): Implications for health, *Molecules*, 22(8), 2017, 1360.
 6. Ronoh A K, Were G M, Mueni M M. Biofortified Crops Can Alleviate Micronutrient Deficiencies: Review of Evidence from Randomized Feeding Trials, *Vitamins and Minerals*, 6(2), 2017, 1-8.
 7. World Health Organization (WHO). Global prevalence of vitamin A deficiency in populations at risk 1995-2005: WHO global database on vitamin A deficiency, Geneva. [Online]: http://apps.who.int/iris/bitstream/10665/44110/1/9789241598019_eng.pdf, [accessed on 30 Dec, 2017].
 8. Darnton-Hill I, Webb P, Harvey P W, Hunt JM, Dalmiya N, Chopra M, Ball M J, Bloem M W, De Benoist B. Micronutrient deficiencies and gender: social and economic costs, *The American journal of clinical nutrition*, 81(5), 2005, 1198S-1205S.
 9. Welch R M, Graham R D. Breeding for micronutrients in staple food crops from a human nutrition perspective, *Journal of experimental botany*, 55(396), 2004, 353-364.
 10. Agarwal A, Khanna P, Baidya D K, Arora M K. Trace elements in critical illness, *Journal of Endocrinology and Metabolism*, 1(2), 2011, 57-63.
 11. Akond A G, Crawford J B, Talukder Z I, Hossain K. Minerals (Zn, Fe, Ca and Mg) and anti-nutrient (phytic acid), *American Journal of Food Technology*, 6(3), 2011, 235-243.
 12. Alzahrani H R, Kumakli H, Ampiah E, Mehari T, Thornton A J, Babyak C M, Fakayode S O. Determination of macro, essential trace elements, toxic heavy metal concentrations, crude oil extracts and ash composition from Saudi Arabian fruits and vegetables having medicinal values, *Arabian Journal of Chemistry*, 10(7), 2017, 906-913.
 13. Kosch M, Hausberg M, Westermann G, Koneke J, Matzkies F, Rahn K H, Kisters K. Alterations in calcium and magnesium content of red cell membranes in patients with primary hypertension, *American journal of hypertension*, 14(3), 2001, 254-258.
 14. Fraga C G. Relevance, essentiality and toxicity of trace elements in human health, *Molecular aspects of medicine*, 26(4-5), 2005, 235-244.
 15. Prasad A S. Discovery of human zinc deficiency: 50 years later, *Journal of Trace Elements in Medicine and Biology*, 26(2-3), 2012, 66-69.
 16. Naismith R T, Shepherd J B, Wehl C C, Tutlam N T, Cross A H. Acute and bilateral blindness due to optic neuropathy associated with copper deficiency, *Archives of neurology*, 66(8), 2009, 1025-1027.
 17. Smith E A, Newland P, Bestwick K G, Ahmed N. Increased whole blood manganese concentrations observed in children with iron deficiency anaemia, *Journal of Trace Elements in Medicine and Biology*, 27(1), 2013, 65-69.
 18. Welch R M. Breeding strategies for biofortified staple plant foods to reduce micronutrient malnutrition globally, *The Journal of nutrition*, 132(3), 2002, 495S-499S.
 19. Nestel P, Bouis H E, Meenakshi J V and Pfeiffer W. Biofortification of staple food

- crops, *The Journal of nutrition*, 136(4), 2009, 1064-1067.
20. Saltzman A, Birol E, Bouis H E, Boy E, De Moura F F, Islam Y, Pfeiffer W H. Biofortification: progress toward a more nourishing future, *Global Food Security*, 2(1), 2013, 9-17.
 21. Graham R D, Welch R M, Bouis H E. Addressing micronutrient malnutrition through enhancing the nutritional quality of staple foods: principles, perspectives and knowledge gaps, *Advances in agronomy*, 70, 2001, 77-142.
 22. Graham R D, Welch R M. Breeding for staple food crops with high micronutrient density, *International Food Policy Research Institute*, 3, 1996, 79.
 23. Moraghan J T, Grafton K. Genetic diversity and mineral composition of common bean seed, *Journal of the Science of Food and Agriculture*, 81(4), 2001, 404-408.
 24. Gouveia C S, Freitas G, de Brito J H, Slaski J J, de Carballo M A P. Nutritional and mineral variability in 52 accessions of common bean varieties (*Phaseolus vulgaris* L.) from Madeira Island, *Agricultural Sciences*, 5(4), 2014, 317-329.
 25. Blair M W, Astudillo C, Grusak M A, Graham R, Beebe S E. Inheritance of seed iron and zinc concentrations in common bean (*Phaseolus vulgaris* L.), *Molecular Breeding*, 23(2), 2009, 197-207.
 26. Hammer O, Harper D A T, Ryan P D. PAST: Paleontological statistics software package for education and data analysis, *Palaeontologia Electronica*, 4(1), 2001, 1-9.
 27. R Core Team. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0. [Online]: <http://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.470.5851&rep=rep1&type=pdf>, [Accessed on 12 March, 2015].
 28. Glowacka A, Klikocka H, Onuch J. Content of zinc and iron in common bean seeds (*Phaseolus vulgaris* L.) in different weed control methods, *Journal of Elementology*, 20(2), 2015, 293-303.
 29. Agung S, McDonald G K. Effects of seed size and maturity on the growth and yield of faba bean (*Vicia faba* L.), *Australian Journal of Agricultural Research*, 49(1), 1998, 79-88.
 30. Mahajan R, Zargar S M, Aezum A M, Farhat S, Gani, M, Agrawal G K and Rakwal R. Evaluation of iron, zinc, and protein contents of common bean (*Phaseolus vulgaris* L.) genotypes: a collection from Jammu and Kashmir, India, *Legume Genomics and Genetics*, 6(2), 2015, 1-7.
 31. Zemolin A E M, Ribeiro N D, Casagrande C R, Silva M J D, Arns F D. Genetic parameters of iron and zinc concentrations in Andean common bean seeds, *Acta Scientiarum. Agronomy*, 38(4), 2016, 439-446.
 32. Possobom M T D F, Ribeiro N D, Domingues L D S, Casagrande C R. Genetic control of iron concentration in Mesoamerican and Andean common bean seeds, *Pesquisa Agropecuaria Brasileira*, 50(5), 2015, 383-391.

Please cite this article in press as: Alex Abaca et al. Determination of micro- and macro-element concentrations and their diversity from the common bean (*phaseolus vulgaris*, l) landraces of Zambia, *International Journal of Nutrition and Agriculture Research*, 5(2), 2018, 46-59.